

New Hampshire Volunteer River Assessment Program 2009 Baker River Water Quality Report



January 2010

**New Hampshire Volunteer River Assessment Program
2009 Baker River Water Quality Report**

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Cover Photo: Baker River, 11-BKR, Warren

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2009 Baker River Volunteers

Larry Cushman

Dick Flanders

Kevin Hopkins

Ellie Murray

John Williams

1.0 INTRODUCTION

1.1. Purpose of Report

Each year the New Hampshire Volunteer River Assessment Program (VRAP) prepares and distributes a water quality report for each volunteer river monitoring group that is based solely on the water quality data collected by that group during a specific year. The reports summarize and interpret the data, particularly as they relate to New Hampshire's surface water quality standards, and serve as a teaching tool and guidance document for future monitoring activities by the individual volunteer groups.

1.2. Report Format

Each report includes the following:

■ Volunteer River Assessment Program Overview

This section includes a description of the history of VRAP, the technical support, training and guidance provided by NHDES, and how data is transmitted to the volunteers and used in surface water quality assessments.

■ Monitoring Program Description

This section provides a description of the volunteer group's monitoring program including monitoring objectives as well as a table and map showing sample station locations.

■ Results and Recommendations

Water quality data collected during the year are summarized on a parameter-by-parameter basis using: (1) a data summary table, which includes the number of samples collected, data ranges, the number of samples meeting New Hampshire water quality standards, and the number of samples adequate for water quality assessments at each station; (2) a discussion of the data; (3) a river graph showing the range of measured values at each station; and (4) a list of applicable recommendations.

Sample results reported as less than the detection limit were assumed equal to one-half the detection limit on the river graphs. This approach simplifies the understanding of the parameter of interest, and specifically helps one to visualize how the river or watershed is functioning from upstream to downstream. In addition, this format allows the reader to better understand potential pollution areas and target those areas for additional sampling or environmental enhancements. Where applicable, the river graph also shows New Hampshire surface water quality standards or levels of concern for comparison purposes.

■ **Appendix A – Water Quality Data**

This appendix includes a spreadsheet detailing the data results and additional information such as data results which do not meet New Hampshire surface water quality standards, and data that is unusable for assessment purposes due to quality control requirements.

■ **Appendix B – Interpreting VRAP Water Quality Parameters**

This appendix provides a brief description of water quality parameters typically sampled by VRAP volunteers and their importance, as well as applicable state water quality criteria or levels of concern.

■ **Appendix C – VRAP Volunteer Monitor Field Sampling Procedures Assessment (*Field Audits*)**

This appendix provides an overview of the VRAP Volunteer Monitor Field Sampling Procedures Assessment (field audit) process with respect to programmatic quality assurance/quality control (QA/QC) guidelines.

2.0 PROGRAM OVERVIEW

2.1 What is VRAP?

In 1998, the New Hampshire Volunteer River Assessment Program was established to promote awareness and education of the importance of maintaining water quality in New Hampshire's rivers and streams. VRAP aims to educate people about river and stream water quality and ecology and to improve water quality monitoring coverage for the protection of water resources.

Today, VRAP loans water quality monitoring equipment, provides technical support, and facilitates educational programs to volunteer groups on numerous rivers and watersheds throughout the state. VRAP volunteers conduct water quality monitoring on an ongoing basis and increase the amount of river water quality information available to local, state and federal governments, which allows for better watershed planning.

2.2 Why is VRAP Important?

VRAP establishes a regular volunteer-driven water sampling program to assist NHDES in evaluating water quality throughout the state. VRAP empowers volunteers with information about the health of New Hampshire's rivers and streams. Regular collection of water quality data allows for early detection of water quality changes allowing NHDES to trace potential problems to their source. Data collected by VRAP volunteers are directly contributing to New Hampshire's obligations under the Clean Water Act. Measurements taken by volunteers are used in assessing the water quality of New Hampshire's river and streams, and are included in reporting to the US Environmental Protection Agency.

2.3 How Does VRAP Work?

VRAP is a cooperative program between NHDES, river groups, local advisory committees, watershed associations, and individuals working to protect New Hampshire's rivers and streams. Volunteers are trained by VRAP staff in the use of water quality monitoring equipment at an annual training workshop. VRAP works with each group to establish monitoring stations and develop a sampling plan.

During the summer months, VRAP receives water quality data from trained volunteers. The data are reviewed for quality assurance, and are entered into the environmental monitoring database at NHDES. During the off-season, VRAP interprets the data and compiles the results into an annual report for each river. VRAP volunteers can use the data as a means of understanding the details of water quality, as well as guide future sampling efforts. NHDES can use the data for making surface water quality assessments, provided that the data met certain quality assurance/quality control guidelines.

2.4 Equipment and Sampling Schedule

VRAP frequently lends and maintains water quality monitoring equipment kits to VRAP groups throughout the state. The kits contain meters and supplies for routine water quality parameter measurements of turbidity, pH, dissolved oxygen, water temperature and specific conductance (conductivity). Other parameters such as nutrients, metals, and *E. coli* can also be studied, although VRAP does not always provide funds to cover laboratory analysis costs. Thus, VRAP encourages groups to pursue other fundraising activities such as association membership fees, special events, in-kind services (non-monetary contributions from individuals and organizations), and grant writing.

Each year, volunteers design and arrange a sampling schedule in cooperation with VRAP staff. Project designs are created through a review and discussion of existing water quality information, such as known and perceived problem areas or locations of exceptional water quality. The interests, priorities, and resources of the partnership determine monitoring locations, parameters, and frequency. VRAP typically recommends sampling every other week from May through September, and VRAP groups are encouraged to organize a long-term sampling program in order to begin to determine trends in river conditions.

2.5 Training and Technical Support

Each VRAP volunteer attends an annual training workshop to receive a demonstration of monitoring protocols and sampling techniques and the calibration and use of water quality monitoring equipment. During the training, volunteers have an opportunity for hands-on use of the equipment and receive instruction in the collection of samples for laboratory analysis.

VRAP groups conduct sampling according to a prearranged monitoring schedule and VRAP protocols. VRAP staff aim to visit each group annually during a scheduled sampling event to verify that volunteers successfully follow the VRAP protocols. If necessary, volunteers are re-trained during the visit, and the group's monitoring coordinator is notified of the result of the verification visit. VRAP groups forward water quality results to NHDES for incorporation into an annual report and state water quality assessment activities.

2.6 Data Usage

Annual Water Quality Reports

Water quality measurements repeated over time create a picture of the fluctuating conditions in rivers and streams and help to determine where improvements, restoration or preservation may benefit the river and the communities it supports. All data collected by volunteers are summarized in water quality reports that are prepared and distributed after the conclusion of the sampling period. VRAP groups can use the reports and data as a means of understanding the details of water quality, guiding future sampling efforts, or determining restoration activities.

New Hampshire Surface Water Quality Assessments

Along with data collected from other water quality programs, specifically the State Ambient River Monitoring Program, applicable volunteer data are used to support periodic NHDES surface water quality assessments. VRAP data are entered into NHDES's environmental monitoring database and are ultimately uploaded to the EPA database. Assessment results and the methodology used to assess surface waters are published by NHDES every two years (i.e., Section 305(b) Water Quality Reports) as required by the federal Clean Water Act. The reader is encouraged to log on to the NHDES web page to review the assessment methodology and list of impaired waters <http://des.nh.gov/organization/divisions/water/wmb/swqa/index.htm/>.

2.7 Quality Assurance/Quality Control

In order for VRAP data to be used in the assessment of New Hampshire's surface waters, the data must meet quality control guidelines as outlined in the VRAP Quality Assurance Project Plan (QAPP). The VRAP QAPP was approved by NHDES and reviewed by EPA in the summer of 2003. The QAPP is reviewed annually and is officially updated and approved every five years. The VRAP quality assurance/quality control measures include a six-step approach to ensuring the accuracy of the equipment and consistency in sampling efforts.

- **Calibration:** Prior to each measurement, the pH and DO meters must be calibrated. Conductivity and turbidity meters are checked against a known standard before the first measurement and after the last one.
- **Replicate Analysis:** A second measurement by each meter is taken from the original sample at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the replicate analysis should be conducted at different stations. Replicates should be measured within 15 minutes of the original measurements.
- **6.0 pH Standard:** A reading of the pH 6.0 buffer is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the 6.0 pH standard check should be conducted at different stations.
- **Zero Oxygen Solution:** A reading of a zero oxygen solution is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the zero oxygen standard check should be conducted at different stations.
- **DI (De-Ionized) Turbidity Blank:** A reading of the DI blank is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the blank check should be conducted at different stations.
- **End of the Day Conductivity and Turbidity Meter Check:** At the conclusion of each sampling day, the conductivity and turbidity meters are re-checked against a known standard.

2.7.1 Measurement Performance Criteria

Precision is calculated for field and laboratory measurements through measurement replicates (instrumental variability) and is calculated for each sampling day. The use of VRAP data for assessment purposes is contingent on compliance with a parameter-specific relative percent difference (RPD) as derived from equation 1, below. Any data exceeding the limits of the individual measures are disqualified from surface water quality assessments. All data that exceeds the limits defined by the VRAP QAPP are acknowledged in the data tables with an explanation of why the data was unusable. Table 1 shows typical parameters studied under VRAP and the associated quality control procedures.

(Equation 1. Relative Percent Difference)

$$RPD = \frac{|x_1 - x_2|}{\frac{x_1 + x_2}{2}} \times 100 \%$$

where x_1 is the original sample and x_2 is the replicate sample

Table 1. Field Analytical Quality Controls

Water Quality Parameter	QC Check	QC Acceptance Limit	Corrective Action	Person Responsible for Corrective Action	Data Quality Indicator
Temperature	Measurement Replicate	RPD < 10% or Absolute Difference < 0.8 C.	Repeat Measurement	Volunteer Monitors	Precision
Dissolved Oxygen	Measurement Replicate	RPD < 10%	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Known Buffer (Zero O ₂ Sol.)	RPD < 10% or Absolute Difference < 0.4 mg/L	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Relative Accuracy
pH	Measurement Replicate	Absolute Difference < 0.3 pH units	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Known Buffer (pH = 6.0)	± 0.1 std units	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Specific Conductance	Measurement Replicate	RPD < 10% or Absolute Difference < 5µS/cm	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Method Blank (Zero Air Reading)	± 5.0 µS/cm	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Turbidity	Measurement Replicate	RPD < 10% or Absolute Difference < 1.0 NTU	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Method Blank (DI Water)	± 0.1 NTU	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Laboratory Parameters	Measurement Replicate	RPD < 20% or Absolute Difference less than ½ the mean value of the parameter in NHDES's Environmental Monitoring Database	Repeat Measurement	Volunteer Monitors	Precision

3.0 METHODS

During the summer of 2001 volunteers from the Baker River Watershed Association began water quality monitoring on the mainstem of the Baker River. The goal of this effort was to provide water quality data from the Baker River relative to surface water quality standards and to allow for the assessment of the river for support of primary contract recreation (swimming). The establishment of a long-term monitoring program will allow for an understanding of the river's dynamics, or variations on a station-by-station and year-to-year basis. The data can also serve as a baseline from which to determine any water pollution problems in the river and/or watershed. The Volunteer River Assessment Program has provided field training, equipment, and technical assistance.

During 2009, trained volunteers from the Baker River VRAP group monitored water quality at 13 stations along the mainstem of the Baker River (Table 3). Stations IDs are designated using a number indicating the relative position of the station and a three-letter code to identify the waterbody name. The higher the station number the more upstream the station is in the watershed. The entire Baker River watershed is designated as Class B waters.

Water quality monitoring was conducted monthly from June through September. Measurements of specific conductance were taken using bottles supplied by NHDES and were analyzed by a hand-held meter supplied by NHDES. Samples for *E.coli* were taken using bottles supplied by NHDES and were stored on ice during transport from the field to lab. Table 2 summarizes the parameters measured, laboratory standard methods, and equipment used.

Table 2. Sampling and Analysis Methods

Parameter	Sample Type	Standard Method	Equipment Used	Laboratory
Specific Conductance	In-Situ	SM 2510	YSI 30	-----
<i>E.coli</i>	Bottle (Sterile)	EPA 1103.1	-----	NHDES

Table 3. Sampling Stations for the Baker River Watershed, NHDES VRAP, 2009

Station ID & AUID	Class	Waterbody Name	Location	Town	Elevation <i>(Rounded to the Nearest 100 Feet)</i>
11-BKR NHRIV700010301-03	B	Baker River	Mooselauke Bridge	Warren	1400
10-BKR NHRIV700010301-03	B	Baker River	Batchelder Bridge	Warren	900
08-BKR NHRIV700010302-14	B	Baker River	Bixby Bridge	Warren	700
07A-BKR NHRIV700010303-09-01	B	Baker River	Town Park	Wentworth	600
07-BKR NHRIV700010303-09-01	B	Baker River	Wentworth Town Beach	Wentworth	600
06A-BKR NHRIV700010303-12	B	Baker River	Silver Bridge	Wentworth	600
05-BKR NHRIV700010305-11	B	Baker River	Sandhill Bridge	Rumney	500
04G-BKR NHRIV700010305-11	B	Baker River	USGS Gauge Station	Rumney	600
04-BKR NHRIV700010305-11	B	Baker River	Stinson Lake Road Bridge/ New Town Bridge	Rumney	500
03-BKR NHRIV700010307-11	B	Baker River	Route 25, Downstream of School Street and Gravel Pit, Upstream of Polar Caves	Rumney	500
02-BKR NHRIV700010307-11	B	Baker River	Covered Bridge/ Smith Lane Road	Plymouth	500
01A-BKR NHRIV700010307-11	B	Baker River	Tenney Mtn. Highway Bridge	Plymouth	500
01-BKR NHRIV700010307-11	B	Baker River	Silver Bridge (Rte 3/25 Bridge)	Plymouth	500

4.0 RESULTS AND RECOMMENDATIONS

Results and recommendations for each monitored parameter are presented in the following sections. For a description of the importance of each parameter and pertinent water quality criteria for these and other parameters, please see Appendix B, “*Interpreting VRAP Water Quality Parameters.*”

4.1 Specific Conductance

Either three or four measurements were taken for specific conductance at 13 stations in the Baker River watershed from Warren to Plymouth (Table 4). Of the 51 measurements taken, 39 met quality assurance/quality control requirements and are usable for New Hampshire’s 2010 surface water quality report to the US Environmental Protection Agency.

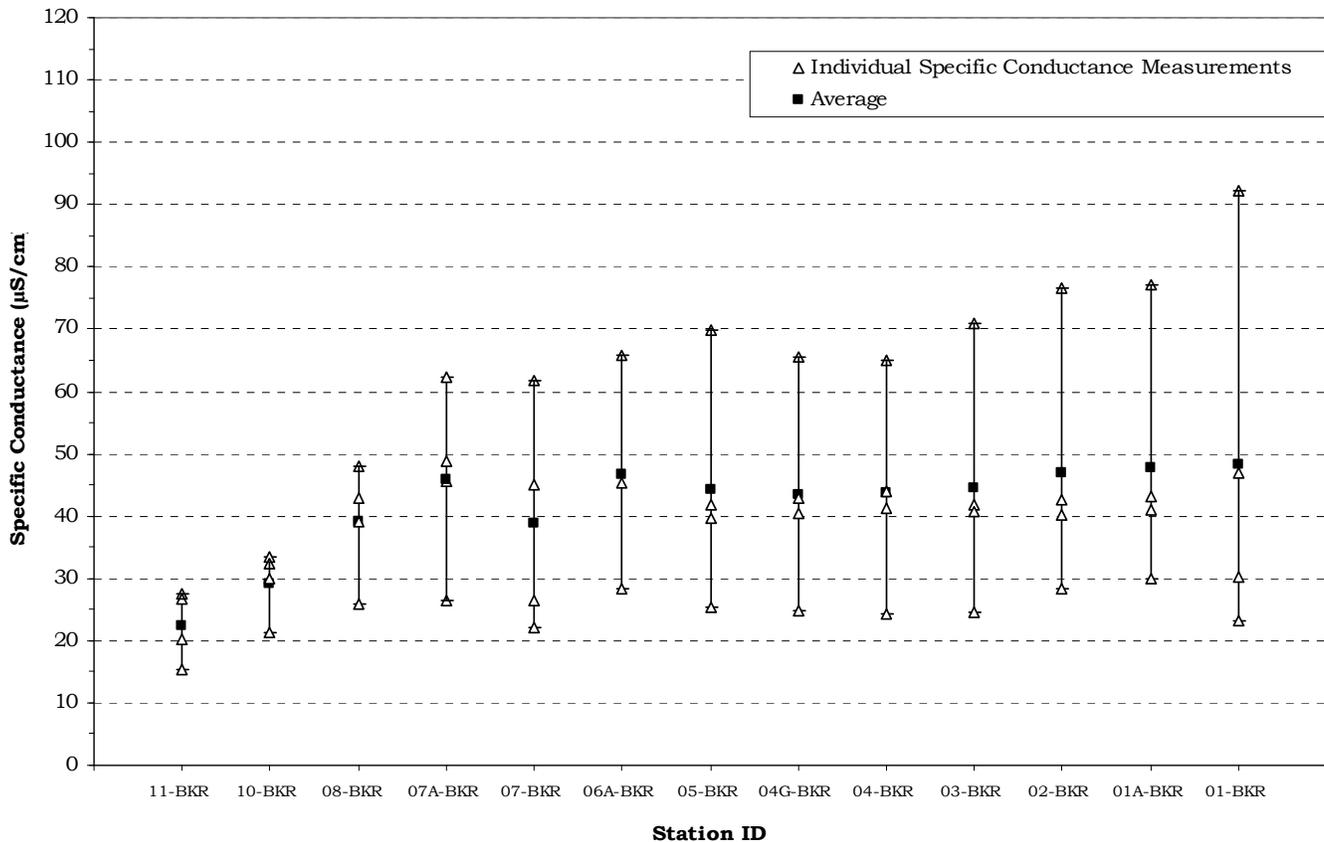
New Hampshire surface water quality standards do not contain numeric criteria for specific conductance although in many fresh surface waters, specific conductance can be used as a surrogate to predict compliance with numeric water quality criteria for chloride.

Table 4. Specific Conductance Data Summary - Baker River, 2009

Station ID	Samples Collected	Data Range (µS/cm)	Acceptable Samples Not Meeting NH Class B Standards (µS/cm as chloride surrogate)	Number of Usable Samples for 2010 NH Surface Water Quality Assessment
11-BKR	4	15.4 - 27.5	0	3
10-BKR	4	21.3 - 33.5	0	3
08-BKR	4	25.8 - 48.1	0	3
07A-BKR	4	26.4 - 62.2	0	3
07-BKR	4	22.1 - 61.8	0	3
06A-BKR	3	28.4 - 65.9	0	3
05-BKR	4	25.3 - 69.9	0	3
04G-BKR	4	24.8 - 65.6	0	3
04-BKR	4	24.4 - 65.0	0	3
03-BKR	4	24.5 - 70.9	0	3
02-BKR	4	28.4 - 76.5	0	3
01A-BKR	4	29.9 - 77.0	0	3
01-BKR	4	23.3 - 92.3	0	3
Total	51	—	0	39

Specific conductance levels were relatively low along the entire reach of the river with the average ranging from 22.5 $\mu\text{S}/\text{cm}$ in the upper portion of the watershed to 48.2 $\mu\text{S}/\text{cm}$ in the lower portion (Figure 1). Higher specific conductance levels can be indicative of pollution from sources such as urban/agricultural runoff, road salt, failed septic systems, or groundwater pollution. Thus, the relatively low specific conductance levels generally indicate low pollutant levels.

**Figure 1. Specific Conductance Statistics for the Baker River Watershed
June 1 - September 8, 2009, NHDES VRAP**



Recommendations

- Continue sampling at all stations in order to develop a long-term data set to better understand trends as time goes on.
- Consider collecting chloride samples at the same time that specific conductance is measured. During the late winter/early spring snowmelt, higher conductivity levels are often seen due to elevated concentrations of chloride in the runoff. Conductivity levels are very closely correlated to chloride levels. Simultaneously measuring chloride and conductivity will allow for a better understanding of their relationship.

4.2 *Escherichia coli*/Bacteria

Four samples were taken for *Escherichia coli* (*E. coli*) at 13 stations in the Baker River watershed from Warren to Plymouth (Table 6). Of the 52 samples taken, 51 met quality assurance/quality control requirements and are usable for New Hampshire's 2010 surface water quality report to the US Environmental Protection Agency.

Class B New Hampshire surface water quality standards for *E.coli* are as follows:

<406 cts/100 ml, based on any single sample or
 ≤126 cts/100 ml, based on a geometric mean calculated from three samples collected within a 60-day period.

Table 5. *E.coli* Data Summary – Baker River, 2009

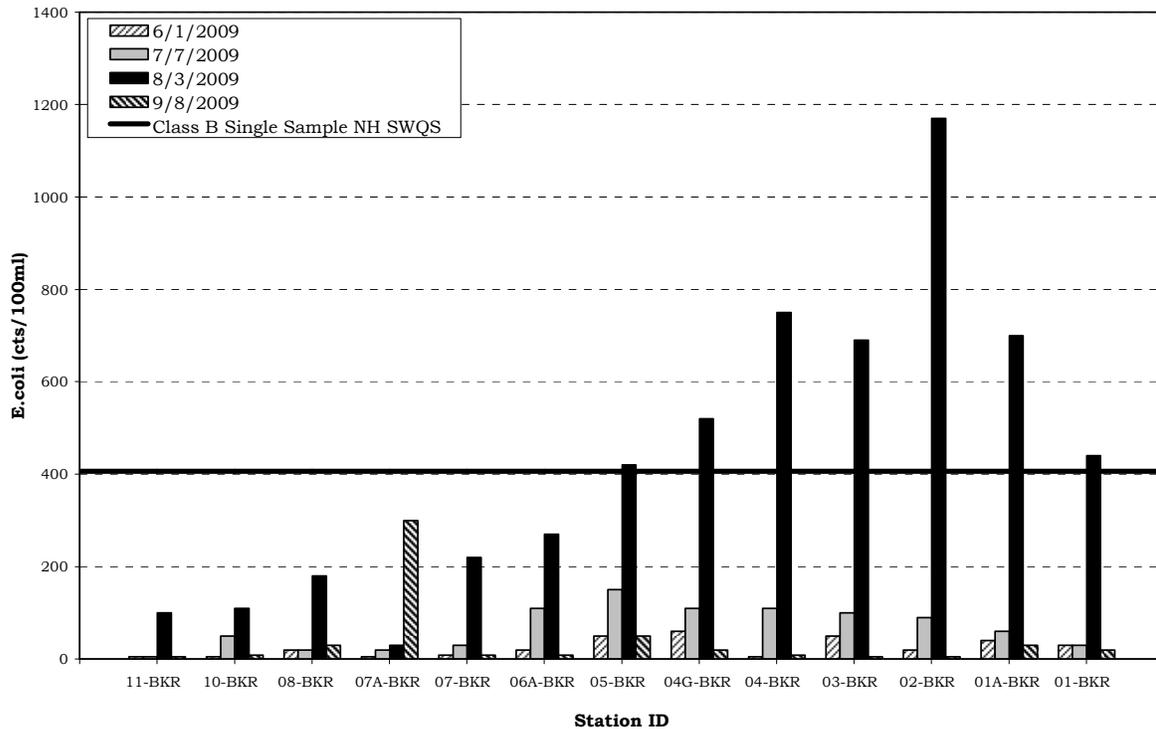
Station ID	Samples Collected	Data Range (cts/100ml)	Acceptable Samples Not Meeting NH Class B Standards	Number of Usable Samples for 2010 NH Surface Water Quality Assessment
11-BKR	4	5 - 100	0	4
10-BKR	4	5 - 110	0	4
08-BKR	4	20 - 180	0	4
07A-BKR	4	5 - 300	0	4
07-BKR	4	9 - 220	0	4
06A-BKR	4	9 - 270	0	4
05-BKR	4	50 - 420	1	4
04G-BKR	4	20 - 520	1	4
04-BKR	4	5 - 750	1	4
03-BKR	4	5 - 690	1	4
02-BKR	4	5 - 1170	1	3
01A-BKR	4	30 - 700	1	4
01-BKR	4	20 - 440	1	4
Total	52	_____	7	51

Seven measurements taken on 8/3/2009 exceeded the New Hampshire Class B surface water quality standard (Figure 3).

In order to fully determine whether a waterbody is meeting surface water standards for *E.coli* a geometric mean must be calculated. A geometric mean is calculated using three samples collected within a 60-day period. During 2009, a geometric mean was not calculated since the samples were collected outside of the 60-day period.

Several factors can contribute to elevated *E. coli* levels, including, but not limited to rain storms, low river flows, the presence of wildlife (e.g., birds), and the presence of septic systems along the river

**Figure 2. *Escherichia coli* Statistics for the Baker River Watershed
June 1 - September 8, 2009, NHDES VRAP**



Recommendations

- In future monitoring collect three samples within 60-day periods during summer to allow for determination of geometric means. Samples should be collected during the critical period of May 24 to September 15. This coincides with the peak contact recreation season.
- Continue to document river conditions and station characteristics (including the presence of wildlife in the area during sampling). At stations with particularly high bacteria levels volunteers can investigate further by moving upstream and taking additional measurements. This will facilitate isolating the location of the cause of the elevated bacteria levels. Those sampling should also look for any potential sources of bacteria such as emission pipes, failed septic systems, farm animals, pet waste, wildlife and waterfowl.

APPENDIX A: 2009 BAKER RIVER WATERSHED VRAP DATA

	Measurements not meeting New Hampshire surface water quality standards
	Measurements not meeting NHDES quality assurance/quality control standards

11-BKR, Baker River, Mooselauke Bridge, Warren

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(uS/cm as chloride surrogate)	<406
06/01/2009	07:15	15.4	<10
07/07/2009	07:20	26.7	<10
08/03/2009	07:15	20.2	100
09/08/2009	07:20	27.5	<10

10-BKR, Baker River, Batchelder Bridge Warren

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(uS/cm as chloride surrogate)	<406
06/01/2009	07:26	33.5	<10
07/07/2009	07:30	29.8	50
08/03/2009	07:25	21.3	110
09/08/2009	07:30	32.4	9

08-BKR, Baker River, Bixby Bridge, Warren

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(uS/cm as chloride surrogate)	<406
06/01/2009	07:37	42.9	20
07/07/2009	07:40	39.1	20
08/03/2009	07:35	25.8	180
09/08/2009	07:40	48.1	30

07A-BKR, Baker River, Town Park, Wentworth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(uS/cm as chloride surrogate)	<406
06/01/2009	07:20	48.9	20
07/07/2009	07:20	45.6	30
08/03/2009	07:10	26.4	300
09/08/2009	07:20	62.2	<10

07-BKR, Baker River, Wentworth Town Beach, Wentworth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:25	22.1	9
07/07/2009	07:30	44.9	30
08/03/2009	07:15	26.4	220
09/08/2009	07:25	61.8	9

06A-BKR, Baker River, Silver Bridge, Saunders Hill Road, Wentworth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:35		20
07/07/2009	07:40	45.3	110
08/03/2009	07:20	28.4	270
09/08/2009	07:30	65.9	9

05-BKR, Baker River, Sandhill Bridge, Rumney

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:04	41.7	50
07/07/2009	07:03	39.6	150
08/03/2009	07:07	25.3	420
09/08/2009	06:58	69.9	50

04G-BKR, Baker River, USGS Gage Station, Rumney

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:43	43.0	60
07/07/2009	07:35	40.5	110
08/03/2009	07:40	24.8	520
09/08/2009	07:30	65.6	20

04-BKR, Baker River, Stinson Hill Road Bridge/New Town Bridge, Rumney

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:15	44.0	5
07/07/2009	07:16	41.3	110
08/03/2009	07:18	24.4	750
09/08/2009	07:07	65.0	9

03-BKR, Baker River, Route 25 D/S of School Street & Gravel Pit, U/S Fron

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:29	41.8	50
07/07/2009	00:00	40.6	100
08/03/2009	07:28	24.5	690
09/08/2009	07:18	70.9	5

02-BKR, Baker River, Covered Bridge/Smith Hill Road, Plymouth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:30	42.7	20
07/07/2009	00:00	40.2	90
08/03/2009	07:12	28.4	1170
09/08/2009	07:12	76.5	<10

01A-BKR, Baker River, Tenney Mountain Highway Bridge, Plymouth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:40	43.2	40
07/07/2009	00:00	41.1	60
08/03/2009	07:25	29.9	700
09/08/2009	07:20	77.0	30

01-BKR, Baker River, Silver Bridge (Route 3/25 Bridge), Plymouth

Date	Time of Sample	Specific Conductance (uS/cm)	<i>E. coli</i> (CTS/100mL)
Standard	NA	(μ S/cm as chloride surrogate)	<406
06/01/2009	07:50	23.3	30
07/07/2009	00:00	46.9	30
08/03/2009	07:35	30.2	440
09/08/2009	07:35	92.3	20

APPENDIX B: Interpreting VRAP Water Quality Monitoring Parameters

Chemical Parameters

Dissolved Oxygen (DO)

- **Unit of Measurement:** concentration in milligrams per liter (mg/L) and percent saturation (%).
- **Description:** A measure of the amount of oxygen in the water: Concentration is a measure of the amount of oxygen in a volume of water; saturation is a measurement of the amount of oxygen in the water compared to the amount of oxygen the water can actually hold at full saturation. Both of these measurements are necessary to accurately determine whether New Hampshire surface water quality standards are met.
- **Importance:** Oxygen is dissolved into the water from the atmosphere, aided by wind and wave action, or by rocky, steep, or uneven stream beds. The presence of dissolved oxygen is vital to bottom-dwelling organisms as well as fish and amphibians. Aquatic plants and algae produce oxygen in the water during the day, and consume oxygen during the night. Bacteria utilize oxygen both day and night when they process organic matter into smaller and smaller particles.

Class A NH Surface Water Quality Standard: 6 mg/L at any place or time, or 75% minimum daily average – (unless naturally occurring).

Class B NH Surface Water Quality Standard: 5 mg/L at any place or time or 75% minimum daily average – (unless naturally occurring).

Several measurements of oxygen saturation taken in a 24-hour period must be averaged to compare to the 75 percent daily average saturation standard. The concentration of dissolved oxygen is dependent on many factors including temperature and sunlight, and tends to fluctuate throughout the day. Saturation values are averaged because a reading taken in the morning may be low due to respiration, while a measurement that afternoon may show that the saturation has recovered to acceptable levels. Water can become saturated with more than 100 percent dissolved oxygen.

pH

- **Unit of Measurement:** units (no abbreviation).
- **Description:** A measure of hydrogen ion activity in water, or, in general terms, the acidity of water. pH is measured on a logarithmic scale of 0 to 14, with 7 being neutral. A high pH indicates alkaline (or basic) conditions and a low pH indicates acidic conditions. pH is influenced by geology and soils, organic acids (decaying leaves and other matter), and human-induced acids from acid rain (which typically has a pH of 3.5 to 5.5).
- **Importance:** pH affects many chemical and biological processes in the water and this is important to the survival and reproduction of fish and other aquatic life. Different organisms flourish within different ranges of pH. Measurements outside of an organism's preferred range can limit growth and reproduction and lead to physiological stress. Low pH can also affect the toxicity of aquatic compounds such as ammonia and certain metals by making them more "available" for uptake by aquatic plants and animals. This can produce conditions that are toxic to aquatic life.

Class A NH Surface Water Quality Standard: Between 6.5 and 8.0 (unless naturally occurring).

Class B NH Surface Water Quality Standard: Between 6.5 and 8.0 (unless naturally occurring).

Sometimes, readings that fall below this range are determined to be naturally occurring. This is often a result of wetlands near the sample station. Wetlands can lower pH because the tannic and humic acids released by decaying plants can cause water to become more acidic.

pH Units	Category
<5.0	High Impact
5.0 – 5.9	Moderate to High Impact
6.0 – 6.4	Normal; Low Impact
6.5 – 8.0	Normal;
6.1 – 8.0	Satisfactory

Specific Conductance or Conductivity

- **Unit of Measurement:** micromhos per centimeter (umhos/cm) or microsiemens per centimeter (uS/cm).
- **Description:** The numerical expression of the ability of water to carry an electrical current at 25° C and a measure of free ion (charged particles) content in the water. These ions can come from natural sources such as bedrock, or human sources such as stormwater runoff. Specific conductance can be used to indicate the presence of chlorides, nitrates, sulfates, phosphates, sodium, magnesium, calcium, iron, and aluminum ions. There is a difference between conductivity and specific conductance. Specific conductance measures the free ion content of water at a *specific* water temperature, whereas conductivity measures the free ion content of water at 25° C. VRAP uses the term “specific conductance” because our conductivity measurements account for temperature. In some studies and programs, the term “conductivity” is used. This term should only be used when the measurement *does not* adjust to a specific temperature.
- **Importance:** Specific conductance readings can help locate potential pollution sources because polluted water usually has a higher specific conductance than unpolluted waters. High specific conductance values often indicate pollution from road salt, septic systems, wastewater treatment plants, or urban/agricultural runoff. Specific conductance can also be related to geology. In unpolluted rivers and streams, geology and groundwater are the primary influences on specific conductance levels.

Class A NH Surface Water Quality Standard: No numeric standard.

Class B NH Surface Water Quality Standard: No numeric standard.

Although there is no formal standard for specific conductance, data collect by VRAP groups and NHDES indicated a very close relationship between specific conductance levels and chloride. In some cases NHDES can use specific conductance measurements as a surrogate for chloride levels. The data collected by NHDES indicate that the chronic chloride standard is correlated with a specific conductance level of approximately 850 uS/cm.

Specific Conductance (uS/cm)	Category
0 – 100	Normal
101 – 200	Low Impact
201 – 500	Moderate Impact
> 501	High Impact
> 850	Likely exceeding chronic chloride standard

Turbidity

- **Unit of Measurement:** Nephelometric Turbidity Units (abbreviated as NTU).
- **Description:** A measurement of the amount of suspended material in the water. This material, which is comprised of particles such as clay, silt, algae, suspended sediment, and decaying plant material, causes light to be scattered and absorbed, rather than transmitted in straight lines through the water.
- **Importance:** Higher turbidity increases water temperatures because suspended particles absorb more heat. This, in turn, reduces dissolved oxygen (DO) concentrations because warm water holds less DO than cold water. Higher turbidity also reduces the amount of light that can penetrate the water, which reduces photosynthesis and DO production. Suspended materials can clog fish gills, reducing disease resistance, lowering growth rates, and affecting egg and larval development. As the particles settle, they can blanket the stream bottom, especially in slower waters, and smother fish eggs and benthic macroinvertebrates. Clean waters are generally associated with low turbidity, but there is a high degree of natural variability involved. Rain events can increase turbidity in surface waters by flushing sediment, organic matter and other materials into the water. Human activities such as vegetation removal and soil disruption can also lead to dramatic increases in turbidity levels.

Class A NH Surface Water Quality Standard: As naturally occurs.

Class B NH Surface Water Quality Standard: Shall not exceed naturally occurring conditions by more than 10 NTU.

Physical Parameters

Temperature

- **Unit of Measurement:** Degrees Celsius (° C)
- **Importance:** Water temperature is a critical parameter for aquatic life and has an impact on other water quality parameters such as dissolved oxygen concentrations, and bacteria activity in water. Water temperature controls the metabolic and reproductive processes of aquatic species and can determine which fish and macroinvertebrate species can survive in a given river or stream.

A number of factors can have an impact on water temperature including the quantity and maturity of riparian vegetation, the rate of flow, the percent of impervious surfaces contributing stormwater, thermal discharges, impoundments and groundwater.

Class A NH Surface Water Quality Standard: No numeric standard; as naturally occurs.

Class B NH Surface Water Quality Standard: No numeric standard

Although there is currently no numerical water quality criteria for water temperature, NHDES is in the process of collecting biological and water temperature data that will contribute to the development of a procedure for assessing rivers and stream based on water temperature and its corresponding impact to the biological integrity of the waterbody.

Chlorophyll-a (Chlor a)

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** An indicator of the biomass, or abundance, of planktonic algae in the river. The technical term “biomass” is used to represent “amount by weight.” Chlorophyll-a can be strongly influenced by phosphorus, which is derived by natural and human activities.

Importance: Because algae is a plant and contains the green pigment chlorophyll-a, the concentration of chlorophyll-a found in the water gives an estimation of the concentration of algae. If the chlorophyll-a concentration increases, this indicates an increase in the algal population.

Class A NH Surface Water Quality Standard: No numeric standard.

Class B NH Surface Water Quality Standard: No numeric standard.

Chlorophyll-a (mg/L)	Category
< 3	Excellent
3 – 7	Good
7 – 15	Less than desirable
> 15	Nuisance

Total Phosphorus (TP)

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** A measure of all forms of phosphorus in the water, including inorganic and organic forms. There are many sources of phosphorus, both natural and human. These include soil and rocks, sewage, animal manure, fertilizer, erosion, and other types of contamination.
- **Importance:** Phosphorus is a nutrient that is essential to plants and animals. However, excess amounts can cause rapid increases in the biological activity in water. Phosphorus is usually the “limiting nutrient” in freshwater streams, which means relatively small amounts can increase algae and chlorophyll-a levels. Algal blooms and/or excessive aquatic plant growth can decrease oxygen levels and make water unattractive. Phosphorus can indicate the presence of septic systems, sewage, animal waste, lawn fertilizer, road and construction erosion, other types of pollution, or natural wetlands and atmospheric deposition.

Class A NH Surface Water Quality Standard: No numeric standard; as naturally occurs.

Class B NH Surface Water Quality Standard: No numeric standard; as naturally occurring, shall contain no phosphorus in such concentrations that would impair any existing or designated uses.

Total Phosphorus (mg/L)	Category
< 0.010	Ideal
0.011 – 0.025	Average
0.026 – 0.050	More than desirable
> 0.051	Excessive (potential nuisance concentration)

Total Kjeldahl Nitrogen (TKN)

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** A measure of the amount of ammonia and organic nitrogen in the water.
- **Importance:** High nitrogen levels can increase algae and chlorophyll-a levels in the river, but is generally less of a concern in fresh water than phosphorus. Nitrogen can indicate the presence of sewage, animal waste, fertilizer, erosion, or other types of pollution.

Class A NH Surface Water Quality Standard: No numeric standard; as naturally occurs.

Class B NH Surface Water Quality Standard: No numeric standard; as naturally occurring, shall contain no nitrogen in such concentrations that would impair any existing or designated uses.

TKN (mg/L)	Category
< 0.25	Ideal
0.26 – 0.40	Average
0.41 – 0.50	More than desirable
> 0.51	Excessive (potential nuisance concentration)

Other Parameters

Chloride

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** The chloride ion (Cl⁻) is found naturally in some surface waters and groundwater. It is also found in high concentrations in seawater. Higher-than-normal chloride concentrations in freshwater is detrimental to water quality. In New Hampshire, applying road salt for winter accident prevention is a large source of chloride to the environment. Unfortunately, this has increased over time due to road expansion and increased vehicle traffic. Road salt (most often sodium chloride) readily dissolves and enters aquatic environments in ionic forms. Although chloride can originate from natural sources, most of the chloride that enters the environment is associated with the storage and application of road salt. As such, chloride-containing compounds commonly enter surface water, soil, and groundwater during late-spring snowmelt (since the ground is frozen during much of the late winter and early spring). Sodium chloride is also used on foods as table salt, and consequently is present in human waste. Thus, sometimes chloride in water can indicate sewage pollution. Saltwater intrusion can also elevate groundwater chlorides in drinking water wells near coastlines. Chloride ions are conservative, which means they are not degraded in the environment and tend to remain in solution, once dissolved. Chloride ions that enter ground water can ultimately be expected to reach surface water and, therefore, influence aquatic environments and humans.
- **Importance:** Research shows elevated chloride levels can be toxic to freshwater aquatic life. Among the species tested, freshwater aquatic plants and invertebrates tend to be the most sensitive to chloride. In order to protect freshwater aquatic life in New Hampshire, the state has adopted acute and chronic chloride criteria.

Acute Standard: 860 mg/L.

Chronic Standard: 230 mg/L.

Escherichia Coliform Bacteria (*E. coli*)

- **Unit of Measurement:** Counts per 100 milliliter (cts/100 mL).
- **Description:** An indicator of the potential presence of pathogens in fresh water. *E. coli* bacteria is a normal component in the large intestines of humans and other warm-blooded animals, and can be excreted in their fecal material. Organisms causing infections or disease (pathogens) are often excreted in the fecal material of humans and other warm-blooded animals.
- **Importance:** *E.coli* bacteria is a good indicator of fecal pollution and the possible presence of pathogenic organisms. In freshwater, *E. coli* concentrations help determine if the water is safe for recreational uses such as swimming.

Several factors can contribute to elevated *E. coli* levels, including, but not limited to rain storms, low river flows, the presence of wildlife, and the presence of septic systems along the river.

Class A NH Surface Water Quality Standard: Unless naturally occurring, shall contain not more than either a geometric mean of 47 *E.coli* cts/100 mL based on at least three samples obtained over a sixty-day period, or greater than 153 *E.coli* cts/100 mL in any one sample.

Class B NH Surface Water Quality Standard: Unless naturally occurring, shall contain not more than either a geometric mean of 126 *E.coli* cts/100 mL based on at least three samples obtained over a sixty-day period, or greater than 406 *E.coli* cts/100 mL in any one sample.

Metals

Depending on the metal concentration, its form (dissolved or particulate), and the hardness of the water, trace metals can be toxic to aquatic life. Metals in dissolved form are generally more toxic than metals in the particulate form. The dissolved metal concentration is dependent on pH, as well as the presence of solids and organic matter that can bind with the metal to render it less toxic.

Hardness is primarily a measure of the calcium and magnesium ion concentrations in water, expressed as calcium carbonate. The hardness concentration affects the toxicity of certain metals. New Hampshire water quality regulations include numeric criteria for a variety of metals. Since dissolved metals are typically found in extremely low concentrations, the potential contamination of samples collected for trace metals analyses has become a primary concern of water quality managers. To prevent such contamination and to ensure reliable results, the use of “clean techniques” is becoming more and more frequent when sampling for dissolved metals. Because of this, sampling for metals may be more costly and require additional effort than in the past.

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2008

APPENDIX C:

2009 VRAP Field Audit

VRAP staff aim to visit each group annually during a scheduled sampling event to verify that volunteers successfully follow the VRAP protocols. If necessary, volunteers are re-trained during the visit, and the group is notified of the result of the verification visit. During the visit, volunteers were assessed in the following five categories:

1) Overall Sampling Procedures

Appropriate storage of meters, sample collection, laboratory sample collection and transportation, beginning and end of day meter checks, collecting a field replicate, performing QA/QC Meter Checks, and ensuring that all calibration and sampling data are properly documented on the VRAP Field Data Sheet and the Laboratory Services Login & Custody Sheet.

2) Turbidity

Inspecting and cleaning of glass turbidity vials prior to measurement of standards and samples, performing the *Initial Turbidity Meter Check*, calibrating the meter to a known standard at the beginning of the sampling day, recording the value of the DI turbidity blank (*QA/QC Meter Check*) once during the sampling day, and performing the “*End of the Day Meter Check*” at the conclusion of the sampling day.

3) pH

Inspecting the pH electrode prior to sampling, calibrating to both pH 7.0 and 4.0 buffers prior to each measurement, rinsing and wiping the pH electrode probe prior to and after the measurement of standards and samples, allowing the pH measurement to stabilize prior to recording the measurement, and recording the value of the 6.0 buffer (*QA/QC Meter Check*) once during the sampling day.

4) Water Temperature/Dissolved Oxygen

Ensuring that the meter is allowed an adequate time to stabilize prior to the first calibration, the meter is calibrated prior to each measurement, the calibration value is properly recorded, the chamber reading is properly recorded, that sufficient time is allowed for readings to stabilize, and that a zero oxygen check (*QA/QC Meter Check*) is completed during the sampling day.

5) Specific Conductance

Performing the *Initial Conductivity Meter Check* using a known standard, allowing for the meter to properly stabilize before recording measurements, properly cleaning the probe between stations, and performing the *End of the Day Meter Check* at the conclusion of the sampling day.

During the field audit, VRAP staff offer important reminders and suggestions to ensure proper sampling techniques and re-trained volunteers in the areas needing improvement. It is important to ensure that all volunteers attend an annual VRAP training workshop prior to the sampling season to familiarize themselves with proper sampling techniques. Please remember to schedule an annual field audit in 2010.