

# **New Hampshire Volunteer River Assessment Program 2010 Nashua River Watershed Water Quality Report**



January 2011



**New Hampshire Volunteer River Assessment Program  
2010 Nashua River Watershed Water Quality Report**

State of New Hampshire  
Department of Environmental Services  
Water Division  
Watershed Management Bureau  
P.O. Box 95  
29 Hazen Drive  
Concord, New Hampshire 03302-0095  
[www.des.nh.gov](http://www.des.nh.gov)

Thomas S. Burack  
Commissioner

Harry T. Stewart  
Water Division Director

**Prepared By:**

Ted Walsh, VRAP Program Manager

Jen Drociak, VRAP Coordinator

Suzanne Richardson, VRAP Assistant

January 2010

# TABLE OF CONTENTS

<b>1. INTRODUCTION.....</b>	<b>3</b>
1.1 Purpose of Report.....	3
1.2 Report Format.....	3
<b>2. PROGRAM OVERVIEW.....</b>	<b>5</b>
2.1 What is VRAP?.....	5
2.2 Why is VRAP Important?.....	5
2.3 How Does VRAP Work?.....	5
2.4 Equipment & Sampling Schedule.....	6
2.5 Training & Technical Support.....	6
2.6 Data Usage.....	6
2.7 Quality Assurance & Quality Control.....	7
<b>3. METHODS.....</b>	<b>9</b>
<b>4. RESULTS &amp; RECOMMENDATIONS.....</b>	<b>11</b>
4.1 Dissolved Oxygen.....	11
4.2 Specific Conductance.....	13
4.3 Water Temperature.....	15
4.4 <i>Eschericia coli</i> /Bacteria.....	17

## **List of Figures and Tables**

Figure 1:	Dissolved Oxygen Concentration Statistics.....	11
Figure 2:	Specific Conductance Statistics.....	14
Figure 3:	Water Temperature Statistics.....	16
Figure 4:	<i>Eschericia coli</i> /Bacteria Statistics.....	18
Table 1:	Field Analytical Quality Controls.....	8
Table 2:	Sampling and Analysis Methods.....	10
Table 3:	Sampling Stations for the Nashua River Watershed, 2010...	10
Table 4:	Dissolved Oxygen Concentration Data Summary.....	11
Table 5:	Specific Conductance Data Summary.....	13
Table 6:	Water Temperature Data Summary.....	15
Table 7:	<i>Eschericia coli</i> /Bacteria Data Summary.....	17

## **List of Appendices**

Appendix A:	2010 Nashua River Watershed Water Quality Data
Appendix B:	Interpreting Water Quality Parameters
Appendix C:	VRAP Volunteer Monitor Field Sampling Procedures Assessment ( <i>Field Audit</i> )

## **ACKNOWLEDGEMENTS**

The New Hampshire Department of Environmental Services Volunteer River Assessment Program extends sincere thanks to the volunteers of the Nashua River Watershed Association for their efforts during 2010. This report was created solely from the data collected by the volunteers listed below. Their time and dedication is an expression of their genuine concern for local water resources and has significantly contributed to our knowledge of river and stream water quality in New Hampshire.

### **2010 Nashua River Watershed Volunteers**

Kathryn Nelson, Water Monitoring Coordinator

Martha Morgan, Water Program Director

Carmen Defillipo, Chief Operator, Pepperell MA Wastewater Treatment Plant

Becky Barth

Beth Chestnutt

Sandy Cofran

Tim Dinsmore

Liz Fletcher

Aron Insinga

Jonas Insinga

Merle Insinga

Audrey Lambert

Tara Lambert

Sara McDougall

Caroline Upham

Steve Wagner

## **1.0 INTRODUCTION**

### **1.1. Purpose of Report**

Each year the New Hampshire Volunteer River Assessment Program prepares and distributes a water quality report for each volunteer river monitoring group that is based solely on the water quality data collected by that group during a specific year. The reports summarize and interpret the data, particularly as they relate to New Hampshire's surface water quality standards, and serve as a teaching tool and guidance document for future monitoring activities by the individual volunteer groups.

### **1.2. Report Format**

Each report includes the following:

#### **■ Volunteer River Assessment Program Overview**

This section includes a description of the history of VRAP, the technical support, training and guidance provided by NHDES, and how data is transmitted to the volunteers and used in surface water quality assessments.

#### **■ Monitoring Program Description**

This section provides a description of the volunteer group's monitoring program including monitoring objectives as well as a table and map showing sample station locations.

#### **■ Results and Recommendations**

Water quality data collected during the year are summarized on a parameter-by-parameter basis using: (1) a data summary table, which includes the number of samples collected, data ranges, the number of samples meeting New Hampshire water quality standards, and the number of samples adequate for water quality assessments at each station; (2) a discussion of the data; (3) a river graph showing the range of measured values at each station; and (4) a list of applicable recommendations.

Sample results reported as less than the detection limit were assumed equal to one-half the detection limit on the river graphs. This approach simplifies the understanding of the parameter of interest, and specifically helps one to visualize how the river or watershed is functioning from upstream to downstream. In addition, this format allows the reader to better understand potential pollution areas and target those areas for additional sampling or environmental enhancements. Where applicable, the river graph also shows New Hampshire surface water quality standards or levels of concern for comparison purposes.

## ■ **Appendix A – Water Quality Data**

This appendix includes a spreadsheet detailing the data results and additional information such as data results which do not meet New Hampshire surface water quality standards, and data that is unusable for assessment purposes due to quality control requirements.

## ■ **Appendix B – Interpreting VRAP Water Quality Parameters**

This appendix provides a brief description of water quality parameters typically sampled by VRAP volunteers and their importance, as well as applicable state water quality criteria or levels of concern.

## ■ **Appendix C – VRAP Volunteer Monitor Field Sampling Procedures Assessment (*Field Audits*)**

This appendix provides an overview of the VRAP Volunteer Monitor Field Sampling Procedures Assessment (field audit) process with respect to programmatic quality assurance/quality control (QA/QC) guidelines.

## **2.0 PROGRAM OVERVIEW**

### **2.1 What is VRAP?**

In 1998, the New Hampshire Volunteer River Assessment Program was established to promote awareness and education of the importance of maintaining water quality in New Hampshire's rivers and streams. VRAP aims to educate people about river and stream water quality and ecology and to improve water quality monitoring coverage for the protection of water resources.

Today, VRAP loans water quality monitoring equipment, provides technical support, and facilitates educational programs to volunteer groups on numerous rivers and watersheds throughout the state. VRAP volunteers conduct water quality monitoring on an ongoing basis and increase the amount of river water quality information available to local, state and federal governments, which allows for better watershed planning.

### **2.2 Why is VRAP Important?**

VRAP establishes a regular volunteer-driven water sampling program to assist NHDES in evaluating water quality throughout the state. VRAP empowers volunteers with information about the health of New Hampshire's rivers and streams. Regular collection of water quality data allows for early detection of water quality changes allowing NHDES to trace potential problems to their source. Data collected by VRAP volunteers are directly contributing to New Hampshire's obligations under the Clean Water Act. Measurements taken by volunteers are used in assessing the water quality of New Hampshire's river and streams, and are included in reporting to the US Environmental Protection Agency.

### **2.3 How Does VRAP Work?**

VRAP is a cooperative program between NHDES, river groups, local advisory committees, watershed associations, and individuals working to protect New Hampshire's rivers and streams. Volunteers are trained by VRAP staff in the use of water quality monitoring equipment at an annual training workshop. VRAP works with each group to establish monitoring stations and develop a sampling plan.

During the summer months, VRAP receives water quality data from trained volunteers. The data are reviewed for quality assurance, and are entered into the environmental monitoring database at NHDES. During the off-season, VRAP interprets the data and compiles the results into an annual report for each river. VRAP volunteers can use the data as a means of understanding the details of water quality, as well as guide future sampling efforts. NHDES can use the data for making surface water quality assessments, provided that the data met certain quality assurance/quality control guidelines.

## **2.4 Equipment and Sampling Schedule**

VRAP frequently lends and maintains water quality monitoring equipment kits to VRAP groups throughout the state. The kits contain meters and supplies for routine water quality parameter measurements of turbidity, pH, dissolved oxygen, water temperature and specific conductance (conductivity). Other parameters such as nutrients, metals, and *E. coli* can also be studied, although VRAP does not always provide funds to cover laboratory analysis costs. Thus, VRAP encourages groups to pursue other fundraising activities such as association membership fees, special events, in-kind services (non-monetary contributions from individuals and organizations), and grant writing.

Each year, volunteers design and arrange a sampling schedule in cooperation with VRAP staff. Project designs are created through a review and discussion of existing water quality information, such as known and perceived problem areas or locations of exceptional water quality. The interests, priorities, and resources of the partnership determine monitoring locations, parameters, and frequency. VRAP typically recommends sampling every other week from May through September, and VRAP groups are encouraged to organize a long-term sampling program in order to begin to determine trends in river conditions.

## **2.5 Training and Technical Support**

Each VRAP volunteer attends an annual training workshop to receive a demonstration of monitoring protocols and sampling techniques and the calibration and use of water quality monitoring equipment. During the training, volunteers have an opportunity for hands-on use of the equipment and receive instruction in the collection of samples for laboratory analysis.

VRAP groups conduct sampling according to a prearranged monitoring schedule and VRAP protocols. VRAP staff aim to visit each group annually during a scheduled sampling event to verify that volunteers successfully follow the VRAP protocols (see Appendix C). If necessary, volunteers are re-trained during the visit, and the group's monitoring coordinator is notified of the result of the verification visit. VRAP groups forward water quality results to NHDES for incorporation into an annual report and state water quality assessment activities.

## **2.6 Data Usage**

### **Annual Water Quality Reports**

Water quality measurements repeated over time create a picture of the fluctuating conditions in rivers and streams and help to determine where improvements, restoration or preservation may benefit the river and the communities it supports. All data collected by volunteers are summarized in water quality reports that are prepared and distributed after the conclusion of the sampling period. VRAP groups can use the reports and data as a means of understanding the details of water quality, guiding future sampling efforts, or determining restoration activities.

## New Hampshire Surface Water Quality Assessments

Along with data collected from other water quality programs, specifically the State Ambient River Monitoring Program, applicable volunteer data are used to support periodic NHDES surface water quality assessments. VRAP data are entered into NHDES's environmental monitoring database and are ultimately uploaded to the EPA database. Assessment results and the methodology used to assess surface waters are published by NHDES every two years (i.e., Section 305(b) Water Quality Reports) as required by the federal Clean Water Act. The reader is encouraged to log on to the NHDES web page to review the assessment methodology and list of impaired waters <http://des.nh.gov/organization/divisions/water/wmb/swqa/index.htm/>.

### 2.7 Quality Assurance/Quality Control

In order for VRAP data to be used in the assessment of New Hampshire's surface waters, the data must meet quality control guidelines as outlined in the VRAP Quality Assurance Project Plan (QAPP). The VRAP QAPP was approved by NHDES and reviewed by EPA in the summer of 2003. The QAPP is reviewed annually and is officially updated and approved every five years. The VRAP quality assurance/quality control measures include a six-step approach to ensuring the accuracy of the equipment and consistency in sampling efforts.

- **Calibration:** Prior to each measurement, the pH and DO meters must be calibrated. Conductivity and turbidity meters are checked against a known standard before the first measurement and after the last one.
- **Replicate Analysis:** A second measurement by each meter is taken from the original sample at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the replicate analysis should be conducted at different stations. Replicates should be measured within 15 minutes of the original measurements.
- **6.0 pH Standard:** A reading of the pH 6.0 buffer is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the 6.0 pH standard check should be conducted at different stations.
- **Zero Oxygen Solution:** A reading of a zero oxygen solution is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the zero oxygen standard check should be conducted at different stations.
- **DI (De-Ionized) Turbidity Blank:** A reading of the DI blank is recorded at one of the stations during the sampling day. If the same sampling schedule is used throughout the monitoring season, the blank check should be conducted at different stations.
- **End of the Day Conductivity and Turbidity Meter Check:** At the conclusion of each sampling day, the conductivity and turbidity meters are re-checked against a known standard.

### 2.7.1 Measurement Performance Criteria

Precision is calculated for field and laboratory measurements through measurement replicates (instrumental variability) and is calculated for each sampling day. The use of VRAP data for assessment purposes is contingent on compliance with a parameter-specific relative percent difference (RPD) as derived from equation 1, below. Any data exceeding the limits of the individual measures are disqualified from surface water quality assessments. All data that exceeds the limits defined by the VRAP QAPP are acknowledged in the data tables with an explanation of why the data was unusable. Table 1 shows typical parameters studied under VRAP and the associated quality control procedures.

(Equation 1. Relative Percent Difference)

$$RPD = \frac{|x_1 - x_2|}{\frac{x_1 + x_2}{2}} \times 100 \%$$

where  $x_1$  is the original sample and  $x_2$  is the replicate sample

**Table 1. Field Analytical Quality Controls**

Water Quality Parameter	QC Check	QC Acceptance Limit	Corrective Action	Person Responsible for Corrective Action	Data Quality Indicator
Temperature	Measurement Replicate	RPD < 10% or Absolute Difference < 0.8 C.	Repeat Measurement	Volunteer Monitors	Precision
Dissolved Oxygen	Measurement Replicate	RPD < 10%	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Known Buffer (Zero O <sub>2</sub> Sol.)	RPD < 10% or Absolute Difference < 0.4 mg/L	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Relative Accuracy
pH	Measurement Replicate	Absolute Difference < 0.3 pH units	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Known Buffer (pH = 6.0)	± 0.1 std units	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Specific Conductance	Measurement Replicate	RPD < 10% or Absolute Difference < 5µS/cm	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Method Blank Known Buffer	± 20% µS/cm	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Turbidity	Measurement Replicate	RPD < 10% or Absolute Difference < 1.0 NTU	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Precision
	Method Blank (DI Water)	± 0.1 NTU	Recalibrate Instrument, Repeat Measurement	Volunteer Monitors	Accuracy
Laboratory Parameters	Measurement Replicate	RPD < 20% or Absolute Difference less than ½ the mean value of the parameter in NHDES's Environmental Monitoring Database	Repeat Measurement	Volunteer Monitors	Precision

### 3.0 METHODS

During the summer of 1993, volunteers from the Nashua River Watershed Association began water quality monitoring on the mainstem and tributaries of the Nashua River. The goal of this effort is to provide water quality data from the Nashua River and its tributaries relative to surface water quality standards and to allow for an assessment of the ecological health of the Nashua River and its tributaries. The continuation of a long-term monitoring program allows for an understanding of the river's dynamics, or variations on a station-by-station and year-to-year basis. The data can also serve as a baseline from which to determine any water pollution problems in the river and/or watershed. The Volunteer River Assessment Program has provided technical assistance and consultation in the development of a Quality Assurance Project Plan (QAPP).

During 2010, trained volunteers from the NRWA monitored water quality at six stations in the Nashua River watershed in New Hampshire (Figure 1, Table 2). Stations IDs are designated using a number indicating the relative position of the station and a three-letter code to identify the waterbody name. The higher the station number the more upstream the station is in the watershed. All stations monitored in 2010 are designated as Class B waters. This class is used to apply the appropriate water quality standard.

Water quality monitoring was conducted monthly from April through October. The NRWA partners with two laboratories - the Devens Wastewater Treatment Facility operated by Earth Tech and the Pepperell Wastewater Treatment Plant. Lab volunteers, lab staff, and the NRWA staff analyze the samples dissolved oxygen, specific conductance, water temperature, and *E.coli*. water temperature and habitat characteristics are recorded by volunteers in the field. In both the field and the lab, quality control procedures are followed according to the NRWA water quality monitoring QAPP. Table 3 summarizes the parameters measured, laboratory standard methods, and equipment used.

**Table 2. Sampling Stations for the Nashua River Watershed, NHDES VRAP, 2008**

<b>Station ID &amp; AUID</b>	<b>Class</b>	<b>Waterbody Name</b>	<b>Location</b>	<b>Town</b>	<b>Elevation (Rounded to the Nearest 100 Feet)</b>
<b>00T-WKR</b> NHRIV700040301-04	<b>B</b>	<b>Walker Brook</b>	Walker Brook - Mason Branch, Route 31	Mason	200
<b>00C-WKG</b> NHRIV700040301-05	<b>B</b>	<b>Walker Brook</b>	Walker Brook - Greenville Branch, Route 31	Mason	200
<b>05-NIS</b> NHRIV700040401-20	<b>B</b>	<b>Nissitissit River</b>	Behind Fire Station on Bond St	Brookline	200
<b>06-NSH</b> NHRIV700040402-05	<b>B</b>	<b>Nashua River</b>	Upstream of Rte 111 Bridge	Hollis	200
<b>04A-NSH</b> NHIMP700040402-03	<b>B</b>	<b>Nashua River</b>	Boat Launch at Mine Falls Park	Nashua	100
<b>02A-NSH</b> NHRIV700040402-09	<b>B</b>	<b>Nashua River</b>	Canal St. at BAE parking lot	Nashua	100

**Table 3. Sampling and Analysis Methods**

<b>Parameter</b>	<b>Sample Type</b>	<b>Standard Method</b>	<b>Equipment Used</b>	<b>Laboratory</b>
Temperature	In Situ	SM 2550	Field Thermometer	-----
Dissolved Oxygen	Laboratory Sample (Bottle)	SM 4500 OC	Winkler Titration with Digital Titrator	Pepperell or Devens WWTF
Specific Conductance	In Situ	SM 2510	Hanna Conductivity Tester	-----
<i>E.coli</i>	Bottle (Sterile)	SM 9223-B	Colilert 24 Testing System	Pepperell or Devens WWTF

## 4.0 RESULTS AND RECOMMENDATIONS

Results and recommendations for each monitored parameter are presented in the following sections. For a description of the importance of each parameter and pertinent water quality criteria for these and other parameters, please see Appendix B, “*Interpreting VRAP Water Quality Parameters.*”

### 4.1 Dissolved Oxygen

Between two and seven measurements were taken in the field for dissolved oxygen concentration at six stations in the Nashua River watershed from Mason to Nashua (Table 4). Of the 27 measurements taken, 19 met quality assurance/quality control requirements and are usable for New Hampshire’s 2010 surface water quality report to the US Environmental Protection Agency.

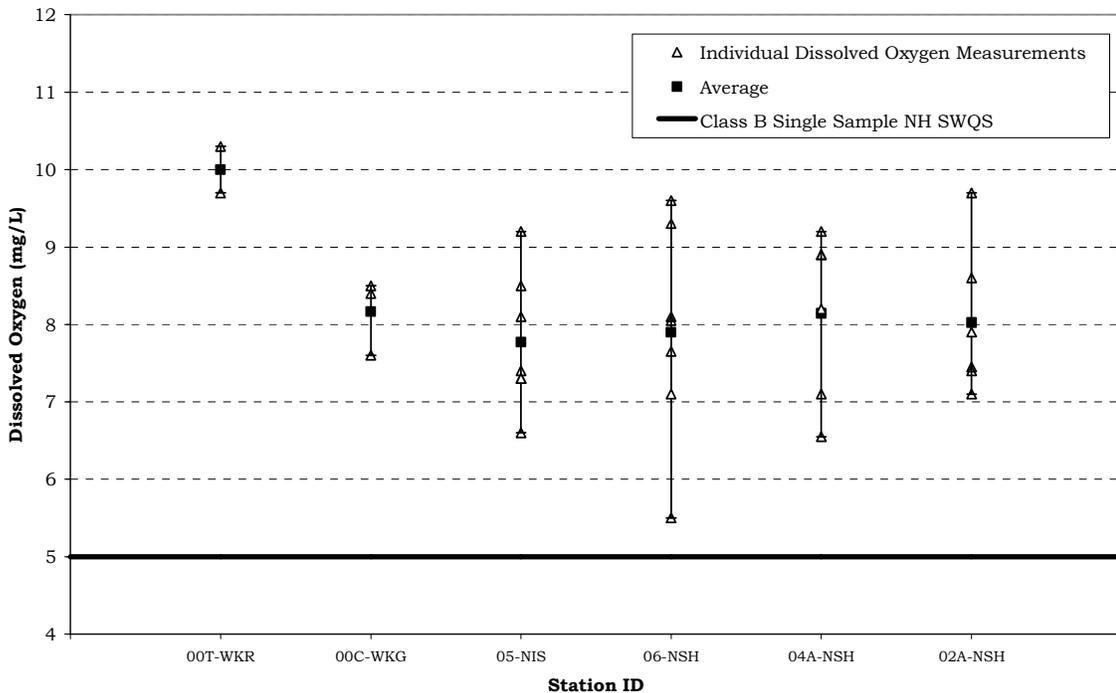
The Class B New Hampshire surface water quality standard for dissolved oxygen includes a minimum concentration of 5.0 mg/L **and** a minimum daily average of 75 percent of saturation. In other words, there are criteria for both concentration and saturation that must be met before the river can be assessed as meeting dissolved oxygen standards. Table 4 reports only dissolved oxygen concentration as more detailed analysis is required to determine if instantaneous dissolved oxygen saturation measurements are above or below water quality standards.

**Table 4. Dissolved Oxygen Concentration (mg/L) Summary – Nashua River Watershed, 2010**

Station ID	Samples Collected	Data Range (mg/L)	Acceptable Samples Not Meeting NH Class B Standards	Number of Usable Samples for 2012 NH Surface Water Quality Assessment
OOT-WKR	2	9.70 - 10.30	0	2
OOC-WKG	3	7.60 - 8.50	0	1
05-NIS	3	6.60 - 9.20	0	1
06-NSH	7	5.50 - 9.60	0	5
04A-NSH	6	6.55 - 9.20	0	5
02A-NSH	6	7.10 - 9.70	0	5
<b>Total</b>	<b>27</b>	_____	<b>0</b>	<b>19</b>

Dissolved oxygen concentration levels met the New Hampshire Class B surface water quality standard at all stations on all occasions. The average ranged from 7.77 mg/L to 10.00 mg/L (Figure 2). Levels of dissolved oxygen sustained above the standards are considered adequate for the support of aquatic life and other desirable water quality conditions.

**Figure 1. Dissolved Oxygen Concentration Statistics for the Nashua River Watershed  
April 17 - October 16, 2010, NHDES VRAP**



## Recommendations

- Continue sampling at all stations in order to develop a long-term data set to better understand trends as time goes on.
- If possible, take measurements between 5 a.m. and 10 a.m., which is when dissolved oxygen is usually the lowest, and between 2 p.m. and 7 p.m. when dissolved oxygen is usually the highest. In general, dissolved oxygen levels are lowest in the early morning when there is low photosynthetic activity and a peak in respiration from organisms throughout the water column. This is the time of least oxygen production and greatest carbon dioxide emission. Peak dissolved oxygen levels occur when photosynthetic activity is at its peak. The greater the amount of photosynthetic activity the greater the production of oxygen as a byproduct of photosynthesis.
- Consider incorporating the use of in-situ dataloggers to automatically record dissolved oxygen saturation levels during a period of several days. The use of these instruments is dependent upon availability, and requires coordination with NHDES.

## 4.2 Specific Conductance

Between four and six measurements were taken in the field for specific conductance at six stations in the Nashua river watershed from Mason to Nashua [Table 5]. Of the 34 measurements taken, all met quality assurance/quality control requirements and are usable for New Hampshire's 2012 surface water quality report to the US Environmental Protection Agency.

Although New Hampshire surface water quality standards do not contain numeric criteria for specific conductance, the New Hampshire Consolidated Assessment and Listing Methodology (CALM) allows for instantaneous specific conductance measurements to be used as a surrogate to predict compliance with numeric water quality criteria for chloride. NHDES has developed a statewide specific conductance to chloride relationship based on simultaneous measurement of specific conductance and chloride.

The Class B New Hampshire surface water quality standard for chloride and corresponding specific conductance measurements are as follows:

Freshwater chronic criterion	230 mg/l	835 uS/cm
Freshwater acute criterion	860 mg/l	2755 uS/cm

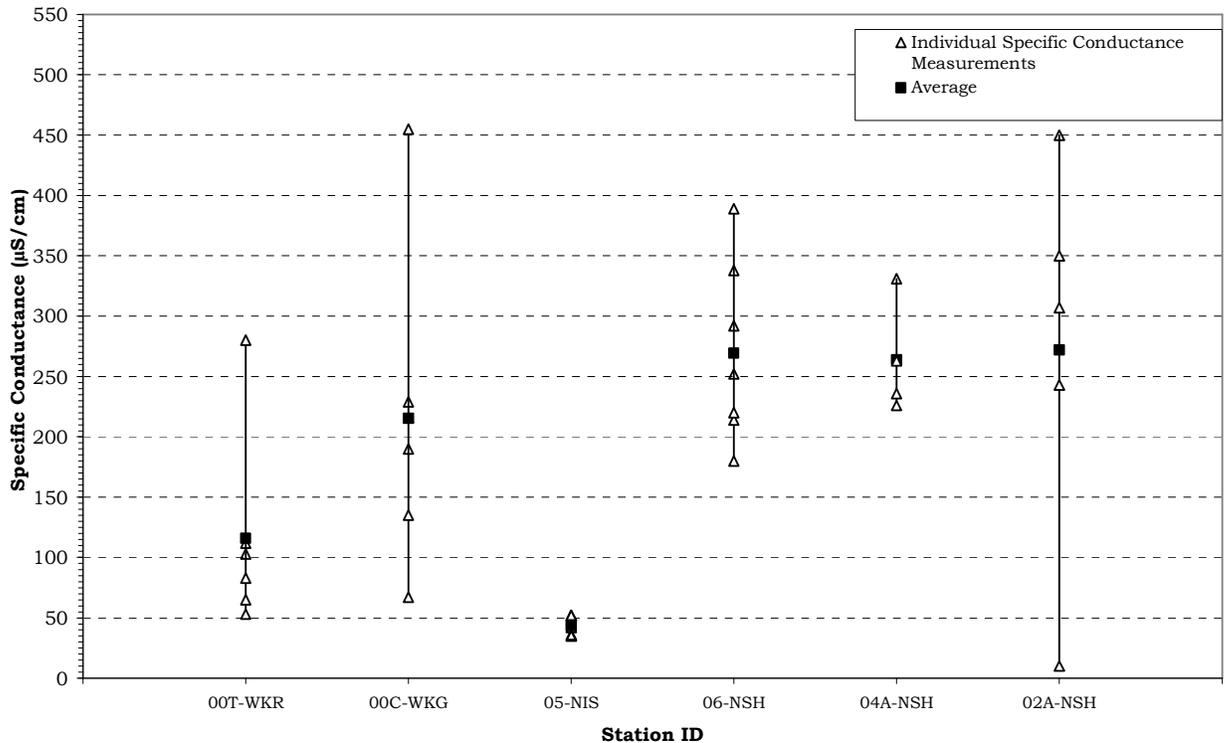
**Table 5. Specific Conductance Data Summary – Nashua River Watershed, 2010**

Station ID	Samples Collected	Data Range (µS/cm)	Acceptable Samples Not Meeting NH Class B Standards (µS/cm as chloride surrogate)	Number of Usable Samples for 2012 NH Surface Water Quality Assessment
<b>00T-WKR</b>	6	53 - 280	0	6
<b>00C-WKG</b>	6	67 - 455	0	6
<b>05-NIS</b>	6	35 - 52	0	6
<b>06-NSH</b>	7	180 - 389	0	7
<b>04A-NSH</b>	4	226 - 331	0	4
<b>02A-NSH</b>	5	10 - 450	0	5
<b>Total</b>	<b>34</b>	—	<b>0</b>	<b>34</b>

Specific conductance levels were variable with the average ranging from 43 µS/cm in the Nissitissit River to 272 µS/cm in the Nashua River (Figure 2).

Higher specific conductance levels can be indicative of pollution from sources such as urban/agricultural runoff, road salt, failed septic systems, or groundwater pollution. Anions (negatively charged elements such as chloride) and cations (positively charged elements such as calcium) are typically found in rivers flowing through more developed areas.

**Figure 2. Specific Conductance Statistics for the Nashua River Watershed  
April 17 -October 16, 2010, NHDES VRAP**



## Recommendations

- Continue sampling at all stations in order to develop a long-term data set to better understand trends as time goes on.
- Consider collecting chloride samples at the same time that specific conductance is measured. During the late winter/early spring snowmelt, higher specific conductance levels are often seen due to elevated concentrations of chloride in the runoff. Specific conductance levels are very closely correlated to chloride levels. Simultaneously measuring chloride and specific conductance will allow for a better understanding of their relationship.
- Continue to incorporate the use of in-situ dataloggers to automatically determine specific conductance levels during rain events, snowmelt, and baseline dry weather conditions. The use of these instruments is dependent upon availability, and requires coordination with NHDES.

### 4.3 Water Temperature

Either six or seven measurements were taken in the field for water temperature at six stations in the Nashua River watershed from Mason to Nashua (Table 6). Of the 37 measurements taken, all met quality assurance/quality control requirements and are usable for New Hampshire's 2010 surface water quality report to the US Environmental Protection Agency.

Although there is currently no numerical water quality criteria for water temperature, NHDES is in the process of collecting biological and water temperature data that will contribute to the development of a procedure for assessing rivers and stream based on water temperature and its corresponding impact to the biological integrity of the waterbody.

**Table 6. Water Temperature Data Summary – Nashua River Watershed, 2008**

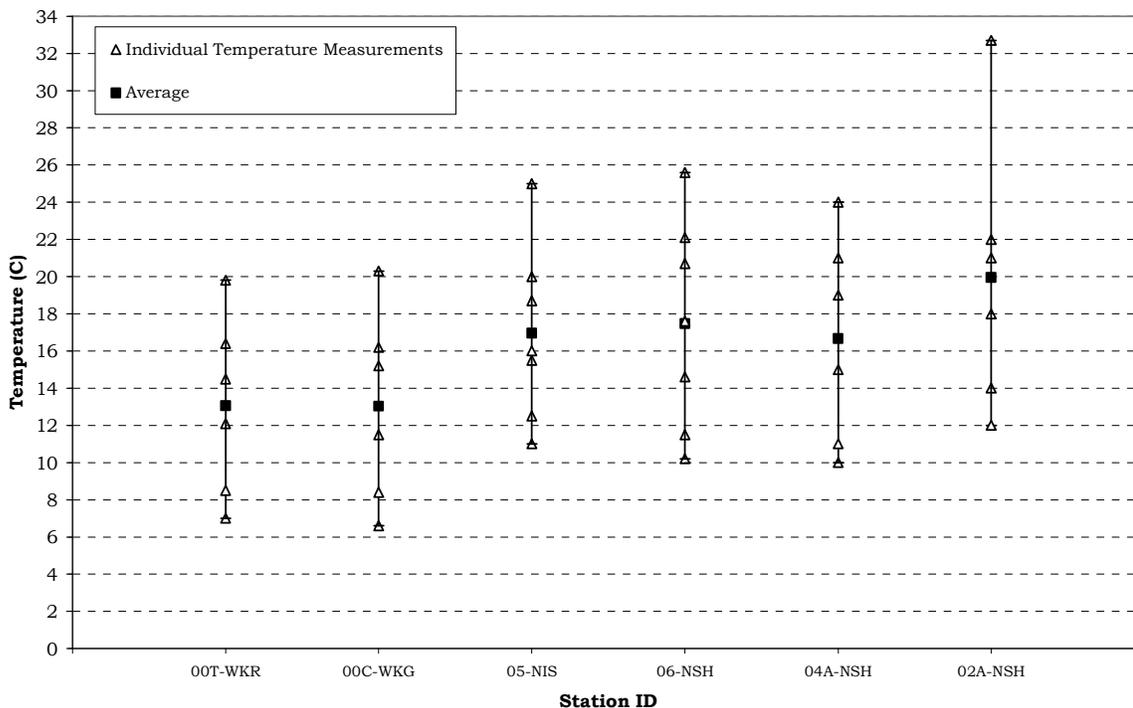
<b>Station ID</b>	<b>Samples Collected</b>	<b>Data Range (°C)</b>	<b>Acceptable Samples Not Meeting NH Class B Standards</b>	<b>Number of Usable Samples for 2010 NH Surface Water Quality Assessment</b>
<b>00T-WKR</b>	6	7.0 - 19.8	Not Applicable	6
<b>00C-WKG</b>	6	6.6 - 20.3	N/A	6
<b>05-NIS</b>	6	11.0 - 25.0	N/A	6
<b>06-NSH</b>	7	10.2 - 25.6	N/A	7
<b>04A-NSH</b>	6	10.0 - 24.0	N/A	6
<b>02A-NSH</b>	6	12.0 - 32.7	N/A	6
<b>Total</b>	<b>37</b>	—	<b>N/A</b>	<b>37</b>

Figure 3 shows the results of instantaneous water temperature measurements taken at six stations in the Nashua River watershed. The average water temperature varied from 13.0 °C. to 20.0 °C.

Water temperature is a critical parameter for aquatic life and has an impact on other water quality parameters such as dissolved oxygen concentrations, and the activity of bacteria in the water. Water temperature controls the metabolic and reproductive processes of aquatic species and can determine which fish and macroinvertebrate species can survive in a given river or stream.

A number of factors can have an impact on water temperature including the quantity and maturity of riparian vegetation along the shoreline, the rate of flow, the percent of impervious surfaces contributing stormwater, thermal discharges, impoundments and the influence of groundwater.

**Figure 3. Water Temperature Statistics for the Nashua River Watershed  
April 17 - October 16, 2010, NHDES VRAP**



### Recommendations

- Continue collecting water temperature data via both instantaneous readings and consider long-term deployment of NHDES water temperature dataloggers.

#### 4.4 *Escherichia coli*/Bacteria

Between five and seven samples were collected for *Escherichia coli* (*E. coli*) at six stations in the Nashua River watershed from Mason to Nashua (Table 8). Of the 35 samples taken, all met quality assurance/quality control requirements and are usable for New Hampshire's 2010 surface water quality report to the US Environmental Protection Agency.

Class B New Hampshire surface water quality standards for *E.coli* are as follows:

- <406 cts/100 ml, based on any single sample, or
- <126 cts/100 ml, based on a geometric mean calculated from three samples collected within a 60-day period.

**Table 8. *E. coli* Data Summary – Nashua River Watershed, 2010**

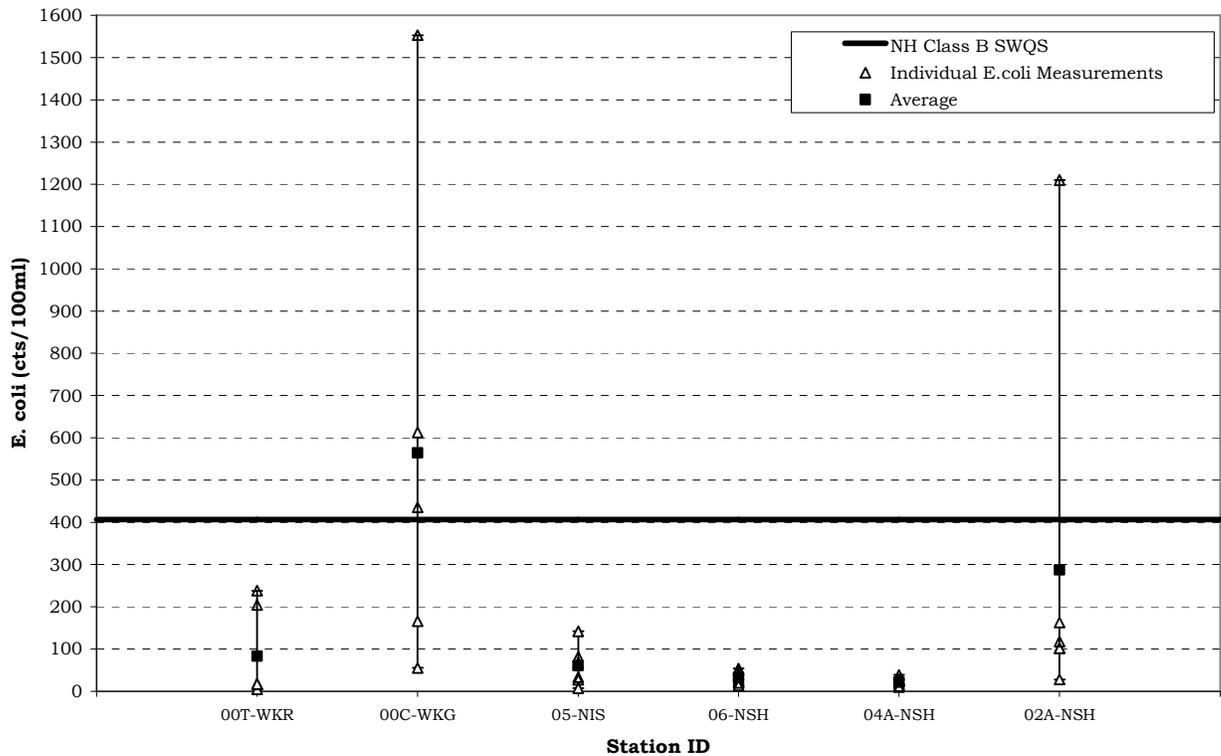
Station ID	Samples Collected	Data Range (cts/100ml)	Acceptable Standards Not Meeting NH Class B Standards	Number of Usable Samples for 2010 NH Surface Water Quality Assessment
00T-WKR	6	5 - 238	0	6
00C-WKG	5	55 - 1553	3	5
05-NIS	5	8 - 142	3	5
06-NSH	7	13 - 54	0	7
04A-NSH	6	9 - 39	0	6
02A-NSH	6	28 - 1210	1	6
<b>Total</b>	<b>35</b>	—	<b>7</b>	<b>35</b>

All but seven measurements met the New Hampshire Class B surface water quality standard for *E.coli* (Figure 4). Stations 00C-WKG, 00C-WKG, and 02A-NSH all had one or more measurement above the standard.

In order to fully determine whether a waterbody is meeting surface water standards for *E.coli* a geometric mean must be calculated. A geometric mean is calculated using three samples collected within a 60-day period. During 2010, a geometric mean was not calculated since the samples were collected outside of the 60-day period.

Several factors can contribute to elevated *E. coli* levels, including, but not limited to rain storms, low river flows, the presence of wildlife, and the presence of septic systems along the river.

**Figure 4. *Escherichia coli* Statistics for the Nashua River Watershed  
April 17 - October 16, 2010, NHDES VRAP**



## Recommendations

- Continue collecting three samples within any 60-day period during the summer to allow for determination of geometric means.
- Continue to document river conditions and station characteristics (including the presence of wildlife in the area during sampling).
- At stations with particularly high bacteria levels volunteers can investigate further by moving upstream and taking additional measurements. This will facilitate isolating the location of the cause of the elevated bacteria levels. Those sampling should also look for any potential sources of bacteria such as emission pipes and failed septic systems.

# APPENDIX A: 2010 NASHUA RIVER WATERSHED VRAP DATA

	Measurements not meeting New Hampshire surface water quality standards
	Measurements not meeting NHDES quality assurance/quality control standards

<sup>A</sup> Specific conductance > 835  $\mu\text{S}/\text{cm}$  indicate exceedance of chronic chloride standard of 230 mg/L

## 00T-WKR, Walker Brook, Mason Branch, Route 31 Mason

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu\text{S}/\text{cm}^{\text{A}}</math></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	7:45	10.30	83	7.0	17
5/15/10	7:55	9.70	65	12.1	5
6/19/10	7:45		53	16.4	16
7/17/10	8:00		103	19.8	205
8/21/10	8:15		280	14.5	17
10/16/10	7:15		112	8.5	238

## 00C-WKG Walker Brook, Greenville Branch, Route 31 Mason

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu\text{S}/\text{cm}^{\text{A}}</math></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	7:46		67	6.6	
5/15/10	7:55		135	11.5	166
6/19/10	7:55	8.50	216	16.2	613
7/17/10	7:45	<del>7.60</del>	229	20.3	1553
8/21/10	8:30		455	15.2	55
10/16/10		<del>8.40</del>	190	8.4	435

## 05-NIS, Nissitissit River, Behind Fire Station on Bond St., Brookline

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu\text{S}/\text{cm}^{\text{A}}</math></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	7:46		67	6.6	
5/15/10	7:55		135	11.5	166
6/19/10	7:55	8.50	216	16.2	613
7/17/10	7:45	<del>7.60</del>	229	20.3	1553
8/21/10	8:30		455	15.2	55
10/16/10		<del>8.40</del>	190	8.4	435

**06-NSH, Nashua River, Upstream of Route 111 Bridge, Hollis**

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu</math>S/cm<sup>A</sup></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	8:20	9.30	180	10.2	23
5/15/10	8:00	9.60	214	14.6	29
6/19/10	8:20	7.65	220	20.7	38
7/17/10	8:20	<del>5.50</del>	338	25.6	48
8/21/10	9:05	8.10	389	22.1	20
9/18/10	8:15	7.10	252	17.6	13
10/16/10	8:35	<del>8.65</del>	292	11.5	54

**04A-NSH, Nashua River, Boat Launch at Mine Falls Park, Nashua**

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu</math>S/cm<sup>A</sup></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	8:18	8.90		10.0	16
5/15/10	8:27	8.90	226	15.0	25
6/19/10	8:18	9.20	236	21.0	12
8/21/10	8:21	6.55	331	24.0	10
9/18/10	7:15	7.10	263	19.0	28
10/16/10	8:15	<del>8.20</del>		11.0	39

**02A-NSH, Nashua River, Canal Street at B.A.E Parking Lot, Nashua**

Date	Time of Sample	DO (mg/L)	Specific Conductance (uS/cm)	Water Temp. (°C)	<i>E. coli</i> (CTS/100mL)
<b>Standard</b>	<b>NA</b>	<b>&gt;5.0</b>	<b>835 <math>\mu</math>S/cm<sup>A</sup></b>	<b>NA</b>	<b>&lt;406</b>
4/17/10	7:45	9.70	10	32.7	28
5/15/10	8:01	8.60	243	14.0	102
6/19/10	7:43	7.40	350	21.0	101
8/21/10	7:45	7.45	450	22.0	1210
9/18/10	7:50	7.10	307	18.0	162
10/16/10	7:45	<del>7.90</del>		12.0	118

# APPENDIX B: Interpreting VRAP Water Quality Monitoring Parameters

## Chemical Parameters

### Dissolved Oxygen (DO)

- **Unit of Measurement:** concentration in milligrams per liter (mg/L) and percent saturation (%).
- **Description:** A measure of the amount of oxygen in the water: Concentration is a measure of the amount of oxygen in a volume of water; saturation is a measurement of the amount of oxygen in the water compared to the amount of oxygen the water can actually hold at full saturation. Both of these measurements are necessary to accurately determine whether New Hampshire surface water quality standards are met.
- **Importance:** Oxygen is dissolved into the water from the atmosphere, aided by wind and wave action, or by rocky, steep, or uneven stream beds. The presence of dissolved oxygen is vital to bottom-dwelling organisms as well as fish and amphibians. Aquatic plants and algae produce oxygen in the water during the day, and consume oxygen during the night. Bacteria utilize oxygen both day and night when they process organic matter into smaller and smaller particles.

**Class A NH Surface Water Quality Standard:** 6 mg/L at any place or time, or 75% minimum daily average – (unless naturally occurring).

**Class B NH Surface Water Quality Standard:** 5 mg/L at any place or time or 75% minimum daily average – (unless naturally occurring).

Several measurements of oxygen saturation taken in a 24-hour period must be averaged to compare to the 75 percent daily average saturation standard. The concentration of dissolved oxygen is dependent on many factors including temperature and sunlight, and tends to fluctuate throughout the day. Saturation values are averaged because a reading taken in the morning may be low due to respiration, while a measurement that afternoon may show that the saturation has recovered to acceptable levels. Water can become saturated with more than 100 percent dissolved oxygen.

### pH

- **Unit of Measurement:** units (no abbreviation).
- **Description:** A measure of hydrogen ion activity in water, or, in general terms, the acidity of water. pH is measured on a logarithmic scale of 0 to 14, with 7 being neutral. A high pH indicates alkaline (or basic) conditions and a low pH indicates acidic conditions. pH is influenced by geology and soils, organic acids (decaying leaves and other matter), and human-induced acids from acid rain (which typically has a pH of 3.5 to 5.5).
- **Importance:** pH affects many chemical and biological processes in the water and this is important to the survival and reproduction of fish and other aquatic life. Different organisms flourish within different ranges of pH. Measurements outside of an organism's preferred range can limit growth and reproduction and lead to physiological stress. Low pH can also affect the toxicity of aquatic compounds such as ammonia and certain metals by making them more "available" for uptake by aquatic plants and animals. This can produce conditions that are toxic to aquatic life.

**Class A/B NH Surface Water Quality Standard:** Between 6.5 and 8.0 (unless naturally occurring).

Sometimes, readings that fall below this range are determined to be naturally occurring. This is often a result of wetlands near the sample station. Wetlands can lower pH because the tannic and humic acids released by decaying plants can cause water to become more acidic.

pH Units	Category
<5.0	High Impact
5.0 – 5.9	Moderate to High Impact
6.0 – 6.4	Normal; Low Impact
6.5 – 8.0	Normal;
6.1 – 8.0	Satisfactory

### **Specific Conductance or Conductivity**

- **Unit of Measurement:** micromhos per centimeter (umhos/cm) or microsiemens per centimeter (uS/cm).
- **Description:** The numerical expression of the ability of water to carry an electrical current at 25° C and a measure of free ion (charged particles) content in the water. These ions can come from natural sources such as bedrock, or human sources such as stormwater runoff. Specific conductance can be used to indicate the presence of chlorides, nitrates, sulfates, phosphates, sodium, magnesium, calcium, iron, and aluminum ions. There is a difference between conductivity and specific conductance. Specific conductance measures the free ion content of water at a *specific* water temperature, whereas conductivity measures the free ion content of water at 25° C. VRAP uses the term “specific conductance” because our conductivity measurements account for temperature. In some studies and programs, the term “conductivity” is used. This term should only be used when the measurement *does not* adjust to a specific temperature.
- **Importance:** Specific conductance readings can help locate potential pollution sources because polluted water usually has a higher specific conductance than unpolluted waters. High specific conductance values often indicate pollution from road salt, septic systems, wastewater treatment plants, or urban/agricultural runoff. Specific conductance can also be related to geology. In unpolluted rivers and streams, geology and groundwater are the primary influences on specific conductance levels.

**Class A/B NH Surface Water Quality Standard:** No numeric standard.

Although NH surface water quality standards do not contain numeric criteria for specific conductance, the NH Consolidated Assessment and Listing Methodology (CALM) allows for instantaneous specific conductance measurements to be used as a surrogate to predict compliance with numeric water quality criteria for chloride. NHDES has developed a statewide specific conductance to chloride relationship based on simultaneous measurement of specific conductance and chloride.

The Class B New Hampshire surface water quality standard for chloride and corresponding specific conductance measurements are as follows:

- Freshwater chronic criterion      230 mg/l                      835 uS/cm
- Freshwater acute criterion          860 mg/l                      2755 uS/cm

Specific Conductance ( <i>uS/cm</i> )	Category
0 – 100	Normal
101 – 200	Low Impact
201 – 500	Moderate Impact
> 501	High Impact
> 835	Exceeding chronic chloride standard

## **Turbidity**

- **Unit of Measurement:** Nephelometric Turbidity Units (abbreviated as NTU).
- **Description:** A measurement of the amount of suspended material in the water. This material, which is comprised of particles such as clay, silt, algae, suspended sediment, and decaying plant material, causes light to be scattered and absorbed, rather than transmitted in straight lines through the water.
- **Importance:** Higher turbidity increases water temperatures because suspended particles absorb more heat. This, in turn, reduces dissolved oxygen (DO) concentrations because warm water holds less DO than cold water. Higher turbidity also reduces the amount of light that can penetrate the water, which reduces photosynthesis and DO production. Suspended materials can clog fish gills, reducing disease resistance, lowering growth rates, and affecting egg and larval development. As the particles settle, they can blanket the stream bottom, especially in slower waters, and smother fish eggs and benthic macroinvertebrates. Clean waters are generally associated with low turbidity, but there is a high degree of natural variability involved. Rain events can increase turbidity in surface waters by flushing sediment, organic matter and other materials into the water. Human activities such as vegetation removal and soil disruption can also lead to dramatic increases in turbidity levels.

**Class A NH Surface Water Quality Standard:** As naturally occurs.

**Class B NH Surface Water Quality Standard:** Shall not exceed naturally occurring conditions by more than 10 NTU.

## **Physical Parameters**

### **Temperature**

- **Unit of Measurement:** Degrees Celsius (° C)
- **Importance:** Water temperature is a critical parameter for aquatic life and has an impact on other water quality parameters such as dissolved oxygen concentrations, and bacteria activity in water. Water temperature controls the metabolic and reproductive processes of aquatic species and can determine which fish and macroinvertebrate species can survive in a given river or stream.

A number of factors can have an impact on water temperature including the quantity and maturity of riparian vegetation, the rate of flow, the percent of impervious surfaces contributing stormwater, thermal discharges, impoundments and groundwater.

**Class A NH Surface Water Quality Standard:** As naturally occurs.

**Class B NH Surface Water Quality Standard:** No numeric standard

Although there is currently no numerical water quality criteria for water temperature, NHDES is in the process of collecting biological and water temperature data that will contribute to the development of a procedure for assessing rivers and stream based on water temperature and its corresponding impact to the biological integrity of the waterbody.

## **Chlorophyll-a (Chlor a)**

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** An indicator of the biomass, or abundance, of planktonic algae in the river. The technical term “biomass” is used to represent “amount by weight.” Chlorophyll-a can be strongly influenced by phosphorus, which is derived by natural and human activities.

**Importance:** Because algae is a plant and contains the green pigment chlorophyll-a, the concentration of chlorophyll-a found in the water gives an estimation of the concentration of algae. If the chlorophyll-a concentration increases, this indicates an increase in the algal population.

**Class A NH Surface Water Quality Standard:** No numeric standard.

**Class B NH Surface Water Quality Standard:** No numeric standard.

<b>Chlorophyll-a (mg/L)</b>	<b>Category</b>
< 3	Excellent
3 – 7	Good
7 – 15	Less than desirable
> 15	Nuisance

## **Total Phosphorus (TP)**

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** A measure of all forms of phosphorus in the water, including inorganic and organic forms. There are many sources of phosphorus, both natural and human. These include soil and rocks, sewage, animal manure, fertilizer, erosion, and other types of contamination.
- **Importance:** Phosphorus is a nutrient that is essential to plants and animals. However, excess amounts can cause rapid increases in the biological activity in water. Phosphorus is usually the “limiting nutrient” in freshwater streams, which means relatively small amounts can increase algae and chlorophyll-a levels. Algal blooms and/or excessive aquatic plant growth can decrease oxygen levels and make water unattractive. Phosphorus can indicate the presence of septic systems, sewage, animal waste, lawn fertilizer, road and construction erosion, other types of pollution, or natural wetlands and atmospheric deposition.

**Class A/B NH Surface Water Quality Standard:** There is no numeric standard for total phosphorus for Class A/B waters. The narrative standard states that “unless naturally occurring, shall contain no phosphorus in such concentrations that would impair any existing or designated uses.” New Hampshire’s surface water regulations (Env-Wq 1700) for Class B waters include narrative criteria for phosphorus which state that “unless naturally occurring, shall contain no phosphorus in such concentrations that would impair any existing or designated uses”. New Hampshire does not currently have numeric nutrient criteria for rivers and streams, but is in the process of developing them. Draft numeric nutrient criteria developed for Vermont and Maine surface waters indicate a maximum allowable summer mean phosphorus level of approximately 0.035 mg/L. Although this value is approximately two to three times typical natural background levels in many rivers and streams, it is considered protective of all designated uses (i.e., swimming, aquatic life, etc). in Vermont and Maine. It’s possible that phosphorus criteria for New Hampshire rivers and streams will be similar.

<b>Total Phosphorus (mg/L)</b>	<b>Category</b>
< 0.010	Ideal
0.011 – 0.025	Average
0.026 – 0.049	More than desirable
≥ 0.050	Potential nuisance concentration

## **Total Kjeldahl Nitrogen (TKN)**

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** A measure of the amount of ammonia and organic nitrogen in the water.
- **Importance:** High nitrogen levels can increase algae and chlorophyll-a levels in the river, but is generally less of a concern in fresh water than phosphorus. Nitrogen can indicate the presence of sewage, animal waste, fertilizer, erosion, or other types of pollution.

**Class A NH Surface Water Quality Standard:** No numeric standard; as naturally occurs.

**Class B NH Surface Water Quality Standard:** No numeric standard; as naturally occurring, shall contain no nitrogen in such concentrations that would impair any existing or designated uses.

<b>TKN (mg/L)</b>	<b>Category</b>
< 0.25	Ideal
0.26 – 0.40	Average
0.41 – 0.50	More than desirable
> 0.51	Excessive (potential nuisance concentration)

## **Other Parameters**

### **Chloride**

- **Unit of Measurement:** Milligrams per liter (mg/L).
- **Description:** The chloride ion (Cl<sup>-</sup>) is found naturally in some surface waters and groundwater. It is also found in high concentrations in seawater. Higher-than-normal chloride concentrations in freshwater is detrimental to water quality. In New Hampshire, applying road salt for winter accident prevention is a large source of chloride to the environment. Unfortunately, this has increased over time due to road expansion and increased vehicle traffic. Road salt (most often sodium chloride) readily dissolves and enters aquatic environments in ionic forms. Although chloride can originate from natural sources, most of the chloride that enters the environment is associated with the storage and application of road salt. As such, chloride-containing compounds commonly enter surface water, soil, and groundwater during late-spring snowmelt (since the ground is frozen during much of the late winter and early spring). Sodium chloride is also used on foods as table salt, and consequently is present in human waste. Thus, sometimes chloride in water can indicate sewage pollution. Saltwater intrusion can also elevate groundwater chlorides in drinking water wells near coastlines. Chloride ions are conservative, which means they are not degraded in the environment and tend to remain in solution, once dissolved. Chloride ions that enter ground water can ultimately be expected to reach surface water and, therefore, influence aquatic environments and humans.
- **Importance:** Research shows elevated chloride levels can be toxic to freshwater aquatic life. Among the species tested, freshwater aquatic plants and invertebrates tend to be the most sensitive to chloride. In order to protect freshwater aquatic life in New Hampshire, the state has adopted acute and chronic chloride criteria.

**Acute Standard:** 860 mg/L.

**Chronic Standard:** 230 mg/L.

## **Escherichia Coliform Bacteria (*E. coli*)**

- **Unit of Measurement:** Counts per 100 milliliter (cts/100 mL).
- **Description:** An indicator of the potential presence of pathogens in fresh water. *E. coli* bacteria is a normal component in the large intestines of humans and other warm-blooded animals, and can be excreted in their fecal material. Organisms causing infections or disease (pathogens) are often excreted in the fecal material of humans and other warm-blooded animals.
- **Importance:** *E.coli* bacteria is a good indicator of fecal pollution and the possible presence of pathogenic organisms. In freshwater, *E. coli* concentrations help determine if the water is safe for recreational uses such as swimming.

Several factors can contribute to elevated *E. coli* levels, including, but not limited to rain storms, low river flows, the presence of wildlife, and the presence of septic systems along the river.

**Class A NH Surface Water Quality Standard:** Unless naturally occurring, shall contain not more than either a geometric mean of 47 *E.coli* cts/100 mL based on at least three samples obtained over a sixty-day period, or greater than 153 *E.coli* cts/100 mL in any one sample.

**Class B NH Surface Water Quality Standard:** Unless naturally occurring, shall contain not more than either a geometric mean of 126 *E.coli* cts/100 mL based on at least three samples obtained over a sixty-day period, or greater than 406 *E.coli* cts/100 mL in any one sample.

## **Metals**

Depending on the metal concentration, its form (dissolved or particulate), and the hardness of the water, trace metals can be toxic to aquatic life. Metals in dissolved form are generally more toxic than metals in the particulate form. The dissolved metal concentration is dependent on pH, as well as the presence of solids and organic matter that can bind with the metal to render it less toxic.

Hardness is primarily a measure of the calcium and magnesium ion concentrations in water, expressed as calcium carbonate. The hardness concentration affects the toxicity of certain metals. New Hampshire water quality regulations include numeric criteria for a variety of metals. Since dissolved metals are typically found in extremely low concentrations, the potential contamination of samples collected for trace metals analyses has become a primary concern of water quality managers. To prevent such contamination and to ensure reliable results, the use of “clean techniques” is becoming more and more frequent when sampling for dissolved metals. Because of this, sampling for metals may be more costly and require additional effort than in the past.

### **New Hampshire Volunteer River Assessment Program**

29 Hazen Drive – PO Box 95  
Concord, NH 03302-0095  
p (603) 271-0699 – f (603) 271-7894  
[www.des.nh.gov](http://www.des.nh.gov)

**2008 (Revised 2010)**

## **APPENDIX C:**

### **2009 VRAP Field Audit**

VRAP staff aim to visit each group annually during a scheduled sampling event to verify that volunteers successfully follow the VRAP protocols. If necessary, volunteers are re-trained during the visit, and the group is notified of the result of the verification visit. During the visit, volunteers were assessed in the following five categories:

#### **1) Overall Sampling Procedures**

Appropriate storage of meters, sample collection, laboratory sample collection and transportation, beginning and end of day meter checks, collecting a field replicate, performing QA/QC Meter Checks, and ensuring that all calibration and sampling data are properly documented on the VRAP Field Data Sheet and the Laboratory Services Login & Custody Sheet.

#### **2) Turbidity**

Inspecting and cleaning of glass turbidity vials prior to measurement of standards and samples, performing the *Initial Turbidity Meter Check*, calibrating the meter to a known standard at the beginning of the sampling day, recording the value of the DI turbidity blank (*QA/QC Meter Check*) once during the sampling day, and performing the “*End of the Day Meter Check*” at the conclusion of the sampling day.

#### **3) pH**

Inspecting the pH electrode prior to sampling, calibrating to both pH 7.0 and 4.0 buffers prior to each measurement, rinsing and wiping the pH electrode probe prior to and after the measurement of standards and samples, allowing the pH measurement to stabilize prior to recording the measurement, and recording the value of the 6.0 buffer (*QA/QC Meter Check*) once during the sampling day.

#### **4) Water Temperature/Dissolved Oxygen**

Ensuring that the meter is allowed an adequate time to stabilize prior to the first calibration, the meter is calibrated prior to each measurement, the calibration value is properly recorded, the chamber reading is properly recorded, that sufficient time is allowed for readings to stabilize, and that a zero oxygen check (*QA/QC Meter Check*) is completed during the sampling day.

#### **5) Specific Conductance**

Performing the *Initial Conductivity Meter Check* using a known standard, allowing for the meter to properly stabilize before recording measurements, properly cleaning the probe between stations, and performing the *End of the Day Meter Check* at the conclusion of the sampling day.

During the field audit, VRAP staff offer important reminders and suggestions to ensure proper sampling techniques and re-trained volunteers in the areas needing improvement. It is important to ensure that all volunteers attend an annual VRAP training workshop prior to the sampling season to familiarize themselves with proper sampling techniques. Please remember to schedule an annual field audit in 2010.